



45 Lyme Road - Suite 304 - Hanover, NH 03755 USA  
Tel: 1-603-643-2325, Fax: 1-603-643-1444

February 23, 2015

Division of Dockets Management (HFA-305)  
Food and Drug Administration  
5630 Fishers Lane, Rm. 1061  
Rockville, MD 20852

**Docket No. FDA-2014-D-1856: Comments to the Draft Guidance Document Titled “Human Cells, Tissues, and Cellular and Tissue-Based Products (HCT/Ps) from Adipose Tissue: Regulatory Considerations; Draft Guidance for Industry” (December 2014)**

Dear Sirs and Madams:

IFATS, the International Federation of Adipose Therapeutics and Sciences, was founded in 2003 by pioneering adipose stem cell biologists and clinician–scientists. Since that time, attendance at the IFATS annual meetings has grown by nearly ten-fold, drawing members from 40 countries in North America, Europe, Africa, the Middle East, Asia, Australia, and Central and South America. The IFATS annual meeting serves as a unique scientific forum that brings together basic scientists, clinicians, translational researchers, and regulatory and biotech representatives to discuss the latest advances in adipose tissue biology and therapeutics. IFATS is formally aligned with the prestigious journal, *Stem Cells*, where a number of the IFATS members serve on the journal’s editorial board, as well as on the editorial board of its sister journal, *Stem Cells Translational Medicine*. Furthermore, in collaboration with the International Society for Cellular Therapy (ISCT), IFATS has provided the scientific community with a detailed description and definition of adipose derived cells (both stromal vascular fraction, or SVF, and adipose-derived stromal/stem cells, or ASCs) in a formal publication in *Cytotherapy*.<sup>1</sup> In addition to including leading basic adipose biologists from around the world, the IFATS membership also includes cardiologists, immunologists, neuroscientists, plastic and reconstructive surgeons, orthopedists, and vascular surgeons who are at the forefront of regenerative medical applications involving adipose tissue and cells. As such, IFATS has the necessary expertise to serve as a resource and think-tank for regulatory agencies examining the safety and efficacy of adipose tissue-related products and therapies. IFATS is committed to patient safety in the translation of new adipose therapies.

IFATS respectfully submits comments to the Draft Guidance Document Titled “Human Cells, Tissues, and Cellular and Tissue-Based Products (HCT/Ps) from Adipose Tissue: Regulatory Considerations; Draft Guidance for Industry” (December 2014).

IFATS requests the FDA to reconsider three main points in the Draft Guidance document. These are 1) the categorization of adipose exclusively and/or primarily as structural; 2) the concept that decellularizing adipose tissue represents more than minimal manipulation; and, 3) the concept that adipose HCT/P's for breast applications would represent non-homologous use.

### **Structural Classification of Adipose Tissue**

The FDA defines HCT/P as “Structural” or “Nonstructural” under 21 CFR 1271.10(a) as:

“4) Either:

- i) The HCT/P does not have a systemic effect and is not dependent upon the metabolic activity of living cells for its primary function; or
- ii) The HCT/P has a systemic effect or is dependent upon the metabolic activity of living cells for its primary function”

We request that the classification of adipose tissue be expanded from exclusively structural to include both structural and/or nonstructural use depending on the intended application. A rigid structural definition would focus solely on adipose tissue *“characteristics for reconstruction, repair, or replacement that relate to its utility to cushion and support the other tissues in the subcutaneous layer (subcutaneum) and skin.”* While this is an important element, there should be equal emphasis on both the structural and nonstructural functions of adipose tissue.

To this point, adipose tissue is a functional unit composed of different cell types, each of which has characteristic nonstructural functions. Cell types include adipocytes, stem and progenitor cells, granulocytes, monocytes, lymphocytes, endothelial cells, pericytes, and stromal cells.<sup>1</sup>

Moreover, adipose is a dynamic tissue in which resident cellular components contribute to nonstructural tissue healing and repair.<sup>2</sup>

We present examples of nonstructural properties of adipose tissue:

### **Adipocytes Appear in Bone Marrow and Have Nonstructural Functions**

The FDA has exempted bone marrow and blood products from regulation under sections 351 and 361. That adipose tissue is present in bone marrow and serves numerous nonstructural functions has been well recognized for over four decades. These nonstructural functions include:<sup>3</sup>

- a. Pre-adipocytes as mesenchymal cells in bone marrow
  - i. Bone marrow contains a spectrum of mesenchymal cells, including pre-adipocytes. When exposed to certain cytokines, pre-adipocytes can differentiate into adipocytes, osteoblasts and chondrocytes depending on the organism’s current needs. This is a nonstructural function.
- b. Bone marrow adipocytes and lympho-hematopoiesis

- i. Pre-adipocytes and adipocytes regulate lympho-hematopoiesis and enable the bone marrow microenvironment to regulate proliferation within blood cell lineages so as to favor erythropoiesis rather than myelopoiesis. This is a nonstructural function.
  - ii. Adipocytes also contain metabolic precursors and energy for the purpose of lympho-hematopoiesis. This physiologic process has nothing to do with providing cushioning and support and therefore is not properly described as a structural use of adipose cells. This is a nonstructural function.
  - iii. Adipocytes contain cholesterol esters, triglycerides and lipoproteins which are essential to the synthesis of plasma membranes during blood cell development. This is a nonstructural function.
- c. Bone marrow and extramedullary adipocytes and energy metabolism:
- i. The energy reserves found in adipocytes assist in homeostatic control of temperature in the bone marrow microenvironment and throughout the body. This is a nonstructural function.
  - ii. Bone marrow and extramedullary adipocytes therefore contribute to the overall energy metabolism of the organism. This is a nonstructural function.
- d. Bone marrow adipose tissue as an endocrine organ: <sup>4</sup>
- i. Bone marrow adipose tissue (MAT) increases during caloric restriction (CR), is responsible for increased adipokine secretion, and alters skeletal muscle adaptation to CR. These and other observations identify MAT as an endocrine organ. This is a nonstructural function.

### **Adipocytes and Adipose Stromal Cells within Adipose Tissue Depots Have Numerous Nonstructural Functions**

We recognize the structural (passive) Roles of adipose tissue, including:

- Insulation (subcutaneous fat)
- Mechanical (infrapatellar fat or Hoffa's fat pad of the knee joint)
- Space occupying (bone marrow fat in the elderly)<sup>5</sup>

However, the multiple nonstructural roles of adipose tissue cannot be ignored. From the first references of fat grafting in the world literature in the late 19<sup>th</sup> century, more than a century ago, surgeons recognized the value of fat for not only providing structure and cushioning, but also for the potential fat has to heal tissues into which it is grafted. In 1893 Gustav Neuber was the first to describe the use of fat grafts. He transplanted fat to the orbital region to heal the

adherent scarring which was the sequela of osteomyelitis. He noted the transformation of facial scarring to more normal appearing skin and subcutaneous tissues.<sup>6</sup>

In 1912, Holländer described the treatment of a breast scar with fat injections. He instructs the reader to sharply release the adhesions between the bone and skin and place fat to prevent the recurrence of the scarring.<sup>7</sup>

In 1926, Charles Conrad Miller<sup>8</sup> developed a new system for injection of fat grafts. Miller described 36 cases of correcting cicatricial contraction on the face and neck with only “moderate shrinkage of the fat. He reported treating with fat grafts “two cases of very persistent parotid fistulas...which defied all other methods of treatment—with excellent results” which he followed for over five years.

Favorable outcomes in the germinal period of fat grafting (1893 – 1926) resulted from fat’s transformational nonstructural uses in addition to its structural uses to provide cushioning and support. Historically and currently, therefore, fat grafting has been used not just for filling or structure, but also for the nonstructural repair of the tissues into which it is placed.<sup>8</sup>

The scientific community has expanded the scope of its understanding of the diverse roles of adipose tissue.<sup>9</sup> A critical factor in shifting the scientific community’s appreciation of the role of adipose tissue was the discovery of the first widely accepted adipokine, leptin, in the mid-1990’s.<sup>10</sup> The realization that adipose tissue secreted proteins with systemic actions on hematopoietic, reproductive, metabolic, and other cells and tissues demonstrated unequivocally that it met the definition of a true “endocrine” organ.<sup>11, 12</sup>

**It is now well-recognized that the many nonstructural roles of adipose tissue include the following:**

#### Endocrine

- Glucose and lipid metabolism and control via adipokine secretion<sup>13</sup>
- Reproductive and endocrine control via adipokine secretion<sup>14-16</sup>
- Immunomodulatory and immunosuppressive systemic control via cytokine and protein factor secretion<sup>17-22</sup>

#### Paracrine

- Angiogenic control via vasculogenic cytokine secretion<sup>22-26</sup>
- Hematopoietic control via cytokine secretion locally and systemically<sup>27</sup>
- Neurogenesis via secretion of cytokine factors<sup>28-34</sup>

#### Hematopoietic potential of adipose stem cells in adipose depots

- Serving as a reservoir for hematopoietic and lymphoid progenitor cells similar to the bone marrow<sup>18, 35, 36</sup>
- Thermogenesis (brown and beige fat)<sup>37-41</sup>

- Energy reservoir (white adipose depots)<sup>42,43</sup>

### Promoting Lactation

- Fat serves as an energy reservoir and nutrient supply for breast epithelial cells. Adipose tissue in the breast undergoes profound changes during pregnancy and parturition in younger females. As pregnancy progresses, the breast epithelium proliferates in a branching manner to occupy the majority of the adjacent adipose tissue and stroma. At parturition, the epithelial cells draw on the lipid reserves of adipocytes within immediate proximity and secrete these nutrients into the milk available to the newborn infant during suckling. As long as the mother continues to breast feed the infant, the epithelial cells remain viable and active; however, if suckling is discontinued for periods of 24 to 48 hours, the epithelial cells undergo rapid apoptosis, leaving pre-adipocytes and adipocytes as the predominant cell within the breast parenchyma. While the presence and organization of epithelial cells within the breast tissue provide it with a unique architecture, the mammary adipocytes themselves show remarkable similarity to adipocytes from elsewhere in the body. Thus, the mammary fat pad displays homology to other adipose tissue depots.<sup>44</sup>

### Regenerative Function

- Fat tissue is a source of local and circulating multipotent progenitor cells capable of repairing and regenerating damaged tissues such as irradiated skin, alleviating fibrotic changes, improving mobility and vitality, and repairing structures such as hair follicles and lymphatics.<sup>45-47</sup>
- Multipotent progenitor cells may be recruited for repair and regeneration of ischemic damage induced by acute myocardial infarction.<sup>48</sup>
- The adipose mesenchymal stem cells also are present in a perivascular position, and serve as progenitors of cells which contribute to vascular network formation and vascular structures.<sup>49-52</sup> As such, the adipose mesenchymal stem cells are located in a position and serve a role shared by mesenchymal stem cells located in nearly all body tissues<sup>53</sup>, and their provision to a range of tissues to enhance vascularity or perfusion constitutes the provision of a cell which is precisely homologous to that already present in the tissue.
- Adipose mesenchymal stem cells induce a monocyte/macrophage phenotype switch from M1 to M2 macrophages, contributing to improved infarct healing postacute myocardial infarction.<sup>54</sup>

## **Additional specific examples of adipose tissue’s nonstructural uses:**

- 1) Modulation of scarring
  - a) Treating old burn scars<sup>55-57</sup>
  - b) Release of adherent scarring/fasciotomies<sup>58</sup>
  - c) Modulation of scarring in primary cleft lip repair<sup>59</sup>
- 2) Reversal of damage caused by therapeutic radiation<sup>60-63</sup>

For BOTH

  - a) Structural (filling tissue defect) uses, and
  - b) Nonstructural tissue repair and regenerative uses<sup>60</sup>
- 3) Treating acute thermal injury<sup>64, 65</sup>
- 4) Treating Pain
  - a) Mitigating implant breast pain<sup>66</sup>
  - b) Improving post-mastectomy pain<sup>67-69</sup>
  - c) Improving lower back pain<sup>70</sup>
  - d) Nerve or neuroma repair<sup>71, 72</sup>
- 5) Healing ulcers
  - a) Treating pressure sores<sup>73</sup>
  - b) Treating chronic non-healing anal fissures and associated stenosis<sup>74</sup>
- 6) Treating vocal fold paralysis<sup>75-77</sup>
- 7) Treating velopharyngeal insufficiency<sup>78</sup>
- 8) Treating scleroderma and systemic sclerosis<sup>79</sup>
- 9) Treating Dupuytren’s disease of the hand<sup>80, 81</sup>
- 10) Treating Raynaud’s phenomenon: After fat grafting, there is improved symptomatology with evidence suggestive of measurably increased perfusion<sup>82</sup>
- 11) Improving tendon repair
  - a) Use of adipose tissue to assist in tenolysis for foot and hand tendon adherence<sup>83</sup>
  - b) Treating adherent tendons and joints in burn patients with fat graft<sup>84</sup>
- 12) Preventing osseous reunion of skull defects<sup>85</sup>
- 13) Improving the quality of skin<sup>86</sup>

## **The Impact of Categorizing Adipose as Exclusively Structural**

Defining all use of adipose tissue as structural despite its many nonstructural uses is particularly problematic in terms of:

1. Defining minimal manipulation
2. Determining homologous use
3. Applying section 351’s “same surgical procedure” exception

## **Minimal Manipulation**

21 CFR 1271.3(f) distinguishes minimal manipulation of structural tissue from minimal manipulation of nonstructural cells and tissues.

- Minimal manipulation of structural tissue consists of processing that does not alter the original relevant characteristics of the tissue relating to the tissue's utility for reconstruction, repair, or replacement.
- For cells and nonstructural tissues that have "a systemic effect or is dependent upon the metabolic activity of living cells for its primary function," minimal manipulation constitutes processing that does not alter "relevant biological characteristics."

Treating all adipose HCT/Ps solely as structural would define minimal manipulation in terms of tissue or cell characteristics relevant to structural properties only. As clearly demonstrated by the many nonstructural uses presented above, applying the concept of minimal manipulation based on cushioning and padding has no relevance when the intended use is nonstructural. Failing to evaluate the characteristics that the FDA has deemed relevant to non-structural use would consequently prevent the proper assessment of risk for application of the tissue. For nonstructural adipose therapy, this could potentially increase risk while simultaneously restricting patient access to therapies.

### **Homologous Use**

The FDA defines homologous use as serving the same basic function in the recipient as in the donor. Thus, to qualify as homologous use under this definition, adipose tissue that serves a nonstructural function in the donor must be used for that same basic purpose in the recipient. The Draft Guidance, however, would again use a structural definition that does not fit nonstructural use. This would preclude all nonstructural uses from qualifying as homologous even though they would otherwise fit FDA's definition of homologous.

### **Same Surgical Procedure Exception**

To qualify for the "same surgical procedure" exception to section 351, the HCT/P must be both minimally manipulated and for homologous use.

As previously explained, subjecting all adipose tissue to the definition of structural tissue would seem to preclude virtually all nonstructural uses from qualifying as minimally manipulated and for homologous use. This would restrict patient access to therapies that are, in fact, minimally manipulated when evaluated in terms of characteristics relevant to intended use, and used for a homologous nonstructural purpose.

## **Decellularized Adipose Tissue and Minimal Manipulation**

Even when adipose tissue is classified as structural, the concept that decellularizing the tissue alters its ability to perform its structural functions and constitutes more than minimal manipulation is unfounded. While adipose tissue is recognized as containing adipocytes, much of the structure of the tissue is imparted by a dense and interconnected framework of fibrous tissue. This fibrous skeleton imparts structural properties irrespective of the presence of cells or lipid<sup>87</sup> and demonstrate notable biomechanical properties of tensile strength and elasticity, both important for padding and cushioning.<sup>88</sup> This collagen skeleton within adipose tissue remains after cells are removed, and multiple reports<sup>89-93</sup> have demonstrated that decellularized adipose tissue retains structural properties and can be injected to impart padding and cushioning of soft tissues. Moreover, the processing of dermis to an acellular form, a well-recognized HCT/P, is comparable to the process of removing cells from adipose tissue. Since decellularization of dermis is regulated under section 361, decellularization of adipose tissue should also be regulated under section 361.

## **Special Considerations for Adipose Tissue and Homologous Use relative to Breast Applications**

In Example B-3 of the Draft Guidance, application of adipose based HCT/Ps to the breast is declared non-homologous use because *“The basic function of breast tissue is to produce milk (lactation) after childbirth. Because this is not a basic function of adipose tissue, using HCT/Ps from adipose tissues for breast augmentation would general be considered a non-homologous use.”* While lactation is a function of the breast, this narrow classification ignores the function of the breast as a secondary sex organ and vital component of a woman’s body image. Indeed, lactation is only utilized in women who have children, and for a limited time span. The important role of the breast as a secondary sex organ is recognized by federal legislation and mandates a woman’s right to breast reconstruction after mastectomy. Importantly, breast reconstruction is often performed in post-menopausal women who will not need to lactate. Additionally, breast reconstruction after mastectomy restores the breast mound but never results in the ability to lactate, and this procedure is commonly performed by transferring adipose tissue flaps. Additionally, fat grafting for breast reconstruction is now a common clinical practice. When considering that the breast is largely composed of fat tissue, applying fat based HCT/Ps to restore breast shape should be clearly considered homologous use.

Importantly, a very common and state-of-the-art method of breast reconstruction involves autologous free tissue flap transfer (free flap breast reconstruction).<sup>94-96</sup> These tissue flaps are completely removed from the body before implanting, and would therefore be considered an HCT/P. By classifying adipose based tissues as non-homologous when applied to the breast, an entire class of Centers for Medicare & Medicaid Services (CMS) approved breast reconstruction procedures would be at risk for not complying with the same surgical procedure exception. Additionally, this would be in opposition to federal legislation that recognizes a woman’s right to breast reconstruction after mastectomy by mandating insurance coverage for the procedures described above.

IFATS wishes to thank the FDA for the opportunity to comment on this draft guidance document. As a multidisciplinary scientific society composed of adipose stem cell biologists and clinician–scientists, we would like the opportunity to engage in dialogue with the FDA. We respectfully request that representatives of the FDA, including the Director of CBER, meet with members of IFATS to further discuss issues surrounding the advancement of adipose based therapies.

### **Respectfully Submitted by IFATS**

J. Peter Rubin, MD, FACS  
IFATS Board Chair and Co-Founder  
UPMC Professor and Chair  
Department of Plastic Surgery  
University of Pittsburgh



Bruce A. Bunnell, PhD  
IFATS President  
Director, Center for Stem Cell Research  
and Regenerative Medicine  
Professor, Department of Pharmacology  
Tulane University School of Medicine



Adam J. Katz, MD, FACS  
IFATS Co-Founder  
Associate Professor  
Director of Plastic Surgery Research,  
Laboratory of BioInnovation and  
Translational Therapeutics  
Division of Plastic Surgery, Department  
of Surgery  
University of Florida College of Medicine



### **IFATS Board of Directors**

Ramon Llull, MD, PhD  
IFATS Co-Founder  
Director of Stem Center  
Courtesy Assistant professor of Plastic  
Surgery  
University of Florida at Gainesville

Pr L. Casteilla, PhD  
Directeur "STROMALab", UMR  
UPS/CNRS5273, EFS, Inserm U1031  
Responsable équipe 1 "Plasticité des  
tissus adipeux"  
BP 84 225  
31 432 Toulouse– France

William Futrell, MD  
IFATS Co-Founder

Sydney R. Coleman, MD  
Adjunct Assistant Professor of Plastic  
Surgery, University of Pittsburgh, and  
New York University

Kacey G. Marra, PhD  
Associate Professor of Plastic Surgery  
Director of Plastic Surgery Research  
University of Pittsburgh School of  
Medicine

Keith March, MD, PhD  
Director, Indiana Center for Vascular  
Biology & Medicine  
Professor of Medicine, Physiology, and  
Biomedical Engineering  
Indiana University School of Medicine

Ricardo L. Rodriguez, MD  
Clinical Instructor, Plastic Surgery  
Johns Hopkins Medical Institutions

Stuart Williams, PhD  
Professor of Physiology and Biophysics  
Director, Bioficial Organs Program  
University of Louisville

Julie Fradette, PhD  
Associate Professor, Dept. of Surgery  
Researcher, Centre de recherche en  
organogénèse expérimentale de  
l'Université Laval / LOEX  
Division of Regenerative Medicine, CHU  
de Québec Research Centre

Jeffrey M. Gimble, MD, PhD  
Adjunct Professor, Center for Stem Cell  
Research & Regenerative Medicine and  
Departments of Medicine, Structural &  
Cellular Biology and Surgery, Tulane  
University School of Medicine  
Chief Scientific Officer, LaCell LLC

Marco N. Helder, PhD  
Department of Orthopedics,  
VU University Medical Center,  
Amsterdam, The Netherlands

Spencer A Brown, PhD  
Director of Research  
Department of Surgery  
Cooper University Hospital

## References

1. Bourin P, Bunnell BA, Casteilla L, Dominici M, Katz AJ, March KL, Redl H, Rubin JP, Yoshimura K, Gimble JM. Stromal cells from the adipose tissue-derived stromal vascular fraction and culture expanded adipose tissue-derived stromal/stem cells: A joint statement of the international federation for adipose therapeutics and science (ifats) and the international society for cellular therapy (isct). *Cytotherapy*. 2013;15:641-648
2. Diaz-Flores L, Gutierrez R, Madrid JF, Varela H, Valladares F, Acosta E, Martin-Vasallo P, Diaz-Flores L, Jr. Pericytes. Morphofunction, interactions and pathology in a quiescent and activated mesenchymal cell niche. *Histol Histopathol*. 2009;24:909-969
3. Gimble JM. The function of adipocytes in the bone marrow stroma. *The New Biologist*. 1990;2:304-312
4. Cawthorn WP, Scheller EL, Learman BS, Parlee SD, Simon BR, Mori H, Ning X, Bree AJ, Schell B, Broome DT. Bone marrow adipose tissue is an endocrine organ that contributes to increased circulating adiponectin during caloric restriction. *Cell metabolism*. 2014;20:368-375
5. Meunier P, Aaron J, Edouard C, VIGNON G. Osteoporosis and the replacement of cell populations of the marrow by adipose tissue: A quantitative study of 84 iliac bone biopsies. *Clinical orthopaedics and related research*. 1971;80:147-154
6. G. N. Über die wiederanheilung vollständig vom körper getrennter, die ganze fettschicht enthaltender hautstücke. *Zbl f Chir* 1893;30:16-17
7. Hollander E, Joseph M. Cosmetic surgery. *Handbuch der Kosmetik*. Leipzig, Germany: Veriag von Velt. 1912;688
8. Miller CC. *Cannula implants and review of implantation technics in esthetic surgery: In two parts*. Oak Press; 1926.
9. Gimble JM FZ. Fat circadian biology. *Journal of applied physiology*. 2009;107:1629-1637
10. Tartaglia LA, Dembski M, Weng X, Deng N, Culpepper J, Devos R, Richards GJ, Campfield LA, Clark FT, Deeds J. Identification and expression cloning of a leptin receptor, ob-r. *Cell*. 1995;83:1263-1271
11. Salgado AJ, Gimble JM. Secretome of mesenchymal stem/stromal cells in regenerative medicine. *Biochimie*. 2013;95:2195
12. Salgado AJ, Reis RL, Sousa N, Gimble JM. Adipose tissue derived stem cells secretome: Soluble factors and their roles in regenerative medicine. *Curr Stem Cell Res Ther*. 2009
13. Khan M, Joseph F. Adipose tissue and adipokines: The association with and application of adipokines in obesity. *Scientifica*. 2014;2014
14. Vicennati V, Garelli S, Rinaldi E, Di Dalmazi G, Pagotto U, Pasquali R. Cross-talk between adipose tissue and the hpa axis in obesity and overt hypercortisolemic states. *Hormone molecular biology and clinical investigation*. 2014;17:63-77
15. Kargi AY, Iacobellis G. Adipose tissue and adrenal glands: Novel pathophysiological mechanisms and clinical applications. *International journal of endocrinology*. 2014;2014
16. Maimoun L, Georgopoulos NA, Sultan C. Endocrine disorders in adolescent and young female athletes: Impact on growth, menstrual cycles, and bone mass acquisition. *The Journal of Clinical Endocrinology & Metabolism*. 2014;99:4037-4050

17. McIntosh K, Zvonic S, Garrett S, Mitchell JB, Floyd ZE, Hammill L, Kloster A, Di Halvorsen Y, Ting JP, Storms RW. The immunogenicity of human adipose-derived cells: Temporal changes in vitro. *Stem cells*. 2006;24:1246-1253
18. McIntosh KR, Frazier T, Rowan BG, Gimble JM. Evolution and future prospects of adipose-derived immunomodulatory cell therapeutics. *Expert review of clinical immunology*. 2013;9:175-184
19. McIntosh KR, Lopez MJ, Borneman JN, Spencer ND, Anderson PA, Gimble JM. Immunogenicity of allogeneic adipose-derived stem cells in a rat spinal fusion model. *Tissue Engineering Part A*. 2009;15:2677-2686
20. Mitchell JB, McIntosh K, Zvonic S, Garrett S, Floyd ZE, Kloster A, Di Halvorsen Y, Storms RW, Goh B, Kilroy G. Immunophenotype of human adipose-derived cells: Temporal changes in stromal-associated and stem cell-associated markers. *Stem cells*. 2006;24:376-385
21. Gimble JM, Dorheim MA, Cheng Q, Medina K, Wang CS, Jones R, Koren E, Pietrangeli C, Kincade PW. Adipogenesis in a murine bone marrow stromal cell line capable of supporting b lineage lymphocyte growth and proliferation: Biochemical and molecular characterization. *European journal of immunology*. 1990;20:379-387
22. Frazier TP, McLachlan JB, Gimble JM, Tucker HA, Rowan BG. Human adipose-derived stromal/stem cells induce functional cd4+ cd25+ foxp3+ cd127- regulatory t cells under low oxygen culture conditions. *Stem cells and development*. 2014;23:968-977
23. Frazier TP, Gimble JM, Kheterpal I, Rowan BG. Impact of low oxygen on the secretome of human adipose-derived stromal/stem cell primary cultures. *Biochimie*. 2013;95:2286-2296
24. Miranville A, Heeschen C, Sengenès C, Curat C, Busse R, Bouloumie A. Improvement of postnatal neovascularization by human adipose tissue-derived stem cells. *Circulation*. 2004;110:349-355
25. Rehman J, Traktuev D, Li J, Merfeld-Clauss S, Temm-Grove CJ, Bovenkerk JE, Pell CL, Johnstone BH, Considine RV, March KL. Secretion of angiogenic and antiapoptotic factors by human adipose stromal cells. *Circulation*. 2004;109:1292-1298
26. Planat-Benard V, Silvestre J-S, Cousin B, André M, Nibbelink M, Tamarat R, Clergue M, Manneville C, Saillan-Barreau C, Duriez M. Plasticity of human adipose lineage cells toward endothelial cells physiological and therapeutic perspectives. *Circulation*. 2004;109:656-663
27. Kilroy GE, Foster SJ, Wu X, Ruiz J, Sherwood S, Heifetz A, Ludlow JW, Stricker DM, Potiny S, Green P, Halvorsen YD, Cheatham B, Storms RW, Gimble JM. Cytokine profile of human adipose-derived stem cells: Expression of angiogenic, hematopoietic, and pro-inflammatory factors. *J Cell Physiol*. 2007;212:702-709
28. Ribeiro CA, Fraga JS, Grãos M, Neves NM, Reis RL, Gimble JM, Sousa N, Salgado AJ. The secretome of stem cells isolated from the adipose tissue and wharton jelly acts differently on central nervous system derived cell populations. *Stem Cell Res Ther*. 2012;3:18
29. Silva NA, Gimble JM, Sousa N, Reis RL, Salgado AJ. Combining adult stem cells and olfactory ensheathing cells: The secretome effect. *Stem cells and development*. 2013;22:1232-1240

30. Cho YJ, Song HS, Bhang S, Lee S, Kang BG, Lee JC, An J, Cha CI, Nam DH, Kim BS. Therapeutic effects of human adipose stem cell-conditioned medium on stroke. *Journal of neuroscience research*. 2012;90:1794-1802
31. Egashira Y, Sugitani S, Suzuki Y, Mishiuro K, Tsuruma K, Shimazawa M, Yoshimura S, Iwama T, Hara H. The conditioned medium of murine and human adipose-derived stem cells exerts neuroprotective effects against experimental stroke model. *Brain research*. 2012;1461:87-95
32. Wei X, Du Z, Zhao L, Feng D, Wei G, He Y, Tan J, Lee WH, Hampel H, Dodel R. Ifats collection: The conditioned media of adipose stromal cells protect against hypoxia-ischemia-induced brain damage in neonatal rats. *Stem Cells*. 2009;27:478-488
33. Wei X, Zhao L, Zhong J, Gu H, Feng D, Johnstone B, March K, Farlow M, Du Y. Adipose stromal cells-secreted neuroprotective media against neuronal apoptosis. *Neuroscience letters*. 2009;462:76-79
34. Zhao L, Wei X, Ma Z, Feng D, Tu P, Johnstone B, March K, Du Y. Adipose stromal cells-conditional medium protected glutamate-induced cngs neuronal death by bdnf. *Neuroscience letters*. 2009;452:238-240
35. Cousin B, André M, Arnaud E, Pénicaud L, Casteilla L. Reconstitution of lethally irradiated mice by cells isolated from adipose tissue. *Biochemical and biophysical research communications*. 2003;301:1016-1022
36. Han J, Koh YJ, Moon HR, Ryoo HG, Cho CH, Kim I, Koh GY. Adipose tissue is an extramedullary reservoir for functional hematopoietic stem and progenitor cells. *Blood*. 2009
37. Harms M, Seale P. Brown and beige fat: Development, function and therapeutic potential. *Nature medicine*. 2013;19:1252-1263
38. Rahman S, Lu Y, Czernik PJ, Rosen CJ, Enerback S, Lecka-Czernik B. Inducible brown adipose tissue, or beige fat, is anabolic for the skeleton. *Endocrinology*. 2013;154:2687-2701
39. Wu J, Cohen P, Spiegelman BM. Adaptive thermogenesis in adipocytes: Is beige the new brown? *Genes & development*. 2013;27:234-250
40. Krings A, Rahman S, Huang S, Lu Y, Czernik P, Lecka-Czernik B. Bone marrow fat has brown adipose tissue characteristics, which are attenuated with aging and diabetes. *Bone*. 2012;50:546-552
41. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND, Schrauwen P, Teule GJ. Cold-activated brown adipose tissue in healthy men. *N Engl J Med*. 2009;360:1500-1508
42. Peirce V, Carobbio S, Vidal-Puig A. The different shades of fat. *Nature*. 2014;510:76-83
43. Enerbäck S, Gimble JM. Lipoprotein lipase gene expression: Physiological regulators at the transcriptional and post-transcriptional level. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*. 1993;1169:107-125
44. Rudolph MC, Neville MC, Anderson SM. Lipid synthesis in lactation: Diet and the fatty acid switch. *Journal of mammary gland biology and neoplasia*. 2007;12:269-281
45. Gimble JM, Katz AJ, Bunnell BA. Adipose-derived stem cells for regenerative medicine. *Circ Res*. 2007;100:1249-1260

46. Bellows CF, Zhang Y, Chen J, Frazier ML, Kolonin MG. Circulation of progenitor cells in obese and lean colorectal cancer patients. *Cancer Epidemiology Biomarkers & Prevention*. 2011;20:2461-2468
47. Bellows CF, Zhang Y, Simmons PJ, Khalsa AS, Kolonin MG. Influence of bmi on level of circulating progenitor cells. *Obesity*. 2011;19:1722-1726
48. Krijnen PA NB, Meinster E, Vo K, Musters RJ, Kamp O, Niessen HW,, Juffermans LJ VDA. Acute myocardial infarction does not affect functional characteristics of adipose derived stem cells in rats, but reduces the number of stem cells in adipose tissue. *IFATS Annual Meeting*. 2014:100
49. Traktuev DO, Merfeld-Clauss S, Li J, Kolonin M, Arap W, Pasqualini R, Johnstone BH, March KL. A population of multipotent cd34-positive adipose stromal cells share pericyte and mesenchymal surface markers, reside in a periendothelial location, and stabilize endothelial networks. *Circulation research*. 2008;102:77-85
50. Traktuev DO, Prater DN, Merfeld-Clauss S, Sanjeevaiah AR, Saadatzadeh MR, Murphy M, Johnstone BH, Ingram DA, March KL. Robust functional vascular network formation in vivo by cooperation of adipose progenitor and endothelial cells. *Circulation research*. 2009;104:1410-1420
51. Merfeld-Clauss S, Gollahalli N, March KL, Traktuev DO. Adipose tissue progenitor cells directly interact with endothelial cells to induce vascular network formation. *Tissue Engineering Part A*. 2010;16:2953-2966
52. Merfeld-Clauss S, Lupov IP, Lu H, Feng D, Compton-Craig P, March KL, Traktuev DO. Adipose stromal cells differentiate along a smooth muscle lineage pathway upon endothelial cell contact via induction of activin a. *Circulation research*. 2014;115:800-809
53. Crisan M, Yap S, Casteilla L, Chen C-W, Corselli M, Park TS, Andriolo G, Sun B, Zheng B, Zhang L. A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell stem cell*. 2008;3:301-313
54. Ter Horst E, Naaijken B, Krijnen P, Van Der Laan A, Piek J, Niessen H. Induction of a monocyte/macrophage phenotype switch by mesenchymal stem cells might contribute to improved infarct healing postacute myocardial infarction. *Minerva cardioangiologica*. 2013;61:617-625
55. Guisantes E, Fontdevila J, Rodríguez G. Autologous fat grafting for correction of unaesthetic scars. *Annals of plastic surgery*. 2012;69:550-554
56. Klinger M, Caviggioli F, Klinger FM, Giannasi S, Bandi V, Banzatti B, Forcellini D, Maione L, Catania B, Vinci V. Autologous fat graft in scar treatment. *Journal of Craniofacial Surgery*. 2013;24:1610-1615
57. Klinger M, Marazzi M, Vigo D, Torre M. Fat injection for cases of severe burn outcomes: A new perspective of scar remodeling and reduction. *Aesthetic plastic surgery*. 2008;32:465-469
58. Khouri RK, Smit JM, Cardoso E, Pallua N, Lantieri L, Mathijssen IM, Khouri Jr RK, Rigotti G. Percutaneous aponeurotomy and lipofilling: A regenerative alternative to flap reconstruction? *Plastic and reconstructive surgery*. 2013;132:1280-1290
59. Balkin DM, Samra S, Steinbacher DM. Immediate fat grafting in primary cleft lip repair. *Journal of Plastic, Reconstructive & Aesthetic Surgery*. 2014;67:1644-1650

60. Rigotti G, Marchi A, Galie M, Baroni G, Benati D, Krampera M, Pasini A, Sbarbati A. Clinical treatment of radiotherapy tissue damage by lipoaspirate transplant: A healing process mediated by adipose-derived adult stem cells. *Plastic and reconstructive surgery*. 2007;119:1409-1422
61. Villani F, Caviggioli F, Klinger F, Klinger M. Rehabilitation of irradiated head and neck tissues by autologous fat transplantation. *Plastic and reconstructive surgery*. 2009;124:2190-2191
62. Chang CC, Thanik VD, Lerman OZ, Saadeh PB, Warren SM, Coleman SR, Hazen A. Treatment of radiation skin damage with coleman fat grafting. *STEM CELLS*. 2007;25:3280-3281
63. Sultan SM, Stern CS, Allen Jr RJ, Thanik VD, Chang CC, Nguyen PD, Canizares O, Szpalski C, Saadeh PB, Warren SM. Human fat grafting alleviates radiation skin damage in a murine model. *Plastic and reconstructive surgery*. 2011;128:363-372
64. Loder S, Peterson JR, Agarwal S, Eboda O, Brownley C, DeLaRosa S, Ranganathan K, Cederna P, Wang SC, Levi B. Wound healing after thermal injury is improved by fat and adipose-derived stem cell isografts. *Journal of Burn Care & Research*. 2015;36:70-76
65. Sultan SM, Barr JS, Butala P, Davidson EH, Weinstein AL, Knobel D, Saadeh PB, Warren SM, Coleman SR, Hazen A. Fat grafting accelerates revascularisation and decreases fibrosis following thermal injury. *Journal of Plastic, Reconstructive & Aesthetic Surgery*. 2012;65:219-227
66. Cuomo R, Zerini I, Botteri G, Barberi L, Nisi G, D'ANIELLO C. Postsurgical pain related to breast implant: Reduction with lipofilling procedure. *In Vivo*. 2014;28:993-996
67. Maione L, Vinci V, Caviggioli F, Klinger F, Banzatti B, Catania B, Lisa A, Klinger M. Autologous fat graft in postmastectomy pain syndrome following breast conservative surgery and radiotherapy. *Aesthetic plastic surgery*. 2014;38:528-532
68. Caviggioli F, Maione L, Forcellini D, Klinger F, Klinger M. Autologous fat graft in postmastectomy pain syndrome. *Plastic and reconstructive surgery*. 2011;128:349-352
69. Caviggioli F, Vinci V, Codolini L. Autologous fat grafting: An innovative solution for the treatment of post-mastectomy pain syndrome. *Breast Cancer*. 2013;20:281-282
70. Salgarello M, Visconti G. The role of sacrolumbar fat grafting in the treatment of spinal fusion instrumentation-related chronic low back pain: A preliminary report. *Spine*. 2014;39:E360-E362
71. Faroni A, Terenghi G, Reid AJ. Adipose-derived stem cells and nerve regeneration: Promises and pitfalls. *Int Rev Neurobiol*. 2013;108:121-136
72. Vaienti L, Gazzola R, Villani F, Parodi PC. Perineural fat grafting in the treatment of painful neuromas. *Techniques in hand & upper extremity surgery*. 2012;16:52-55
73. Marangi GF, Pallara T, Cagli B, Schena E, Giurazza F, Faiella E, Zobel BB, Persichetti P. Treatment of early-stage pressure ulcers by using autologous adipose tissue grafts. *Plastic surgery international*. 2014;2014
74. Lolli P, Malleo G, Rigotti G. Treatment of chronic anal fissures and associated stenosis by autologous adipose tissue transplant: A pilot study. *Diseases of the Colon & Rectum*. 2010;53:460-466

75. Cantarella G, Baracca G, Forti S, Gaffuri M, Mazzola R. Outcomes of structural fat grafting for paralytic and non-paralytic dysphonia. *Acta Otorhinolaryngologica Italica*. 2011;31:154
76. DeFatta RA, DeFatta RJ, Sataloff RT. Laryngeal lipotransfer: Review of a 14-year experience. *Journal of Voice*. 2013;27:512-515
77. Sataloff RT. Autologous fat implantation for vocal fold scar. *Current opinion in otolaryngology & head and neck surgery*. 2010;18:503-506
78. Cantarella G, Mazzola RF, Mantovani M, Baracca G, Pignataro L. Treatment of velopharyngeal insufficiency by pharyngeal and velar fat injections. *Otolaryngology--Head and Neck Surgery*. 2011;145:401-403
79. Papa N, Luca G, Sambataro D, Zaccara E, Maglione W, Gabrielli A, Fraticelli P, Moroncini G, Beretta L, Santaniello A. Regional implantation of autologous adipose tissue-derived cells induces a prompt healing of long-lasting indolent digital ulcers in patients with systemic sclerosis. *Cell transplantation*. 2014
80. Hovius SE, Kan HJ, Smit X, Selles RW, Cardoso E, Khouri RK. Extensive percutaneous aponeurotomy and lipografting: A new treatment for dupuytren disease. *Plastic and reconstructive surgery*. 2011;128:221-228
81. Verhoekx JS, Mudera V, Walbeehm ET, Hovius SE. Adipose-derived stem cells inhibit the contractile myofibroblast in dupuytren's disease. *Plastic and reconstructive surgery*. 2013;132:1139-1148
82. Bank J, Fuller SM, Henry GI, Zachary LS. Fat grafting to the hand in patients with raynaud phenomenon: A novel therapeutic modality. *Plastic and reconstructive surgery*. 2014;133:1109-1118
83. Damgaard OE, Siemssen PA. Lipografted tenolysis. *Journal of Plastic, Reconstructive & Aesthetic Surgery*. 2010;63:e637-e638
84. Colonna M, Scarcella M, d'Alcontres F, Delia G, Lupo F. Should fat graft be recommended in tendon scar treatment? Considerations on three cases (two feet and a severe burned hand). *European review for medical and pharmacological sciences*. 2014;18:753-759
85. Merikanto JE, Alhopuro S, Ritsilä VA. Free fat transplant prevents osseous reunion of skull defects: A new approach in the treatment of craniosynostosis. *Scandinavian Journal of Plastic and Reconstructive Surgery and Hand Surgery*. 1987;21:183-188
86. Mojallal A, Lequeux C, Shipkov C, Breton P, Foyatier J-L, Braye F, Damour O. Improvement of skin quality after fat grafting: Clinical observation and an animal study. *Plastic and reconstructive surgery*. 2009;124:765-774
87. Lockwood TE. Superficial fascial system (sfs) of the trunk and extremities: A new concept. *Plastic and reconstructive surgery*. 1991;87:1009-1018
88. Song AY, Askari M, Azemi E, Alber S, Hurwitz DJ, Marra KG, Shestak KC, Debski R, Rubin JP. Biomechanical properties of the superficial fascial system. *Aesthetic Surgery Journal*. 2006;26:395-403
89. Flynn L. The use of decellularized adipose tissue to provide an inductive microenvironment for the adipogenic differentiation of human adipose-derived stem cells. *Biomaterials*. 2010;31:4715-4724

90. Brown BN, Freund JM, Han L, Rubin JP, Reing JE, Jeffries EM, Wolf MT, Tottey S, Barnes CA, Ratner BD. Comparison of three methods for the derivation of a biologic scaffold composed of adipose tissue extracellular matrix. *Tissue Engineering Part C: Methods*. 2011;17:411-421
91. Wu I, Nahas Z, Kimmerling KA, Rosson GD, Elisseff JH. An injectable adipose matrix for soft tissue reconstruction. *Plastic and reconstructive surgery*. 2012;129:1247
92. Omid E, Fuetterer L, Mousavi SR, Armstrong RC, Flynn LE, Samani A. Characterization and assessment of hyperelastic and elastic properties of decellularized human adipose tissues. *Journal of biomechanics*. 2014;47:3657-3663
93. Wang L, Johnson JA, Zhang Q, Beahm EK. Combining decellularized human adipose tissue extracellular matrix and adipose-derived stem cells for adipose tissue engineering. *Acta biomaterialia*. 2013;9:8921-8931
94. Healy C, Allen Sr RJ. The evolution of perforator flap breast reconstruction: Twenty years after the first diep flap. *Journal of reconstructive microsurgery*. 2014;30:121-125
95. LoTempio MM, Allen RJ. Breast reconstruction with sgap and igap flaps. *Plastic and reconstructive surgery*. 2010;126:393-401
96. Erić M, Mihić N, Krivokuća D. Breast reconstruction following mastectomy; patient's satisfaction. *Acta Chir Belg*. 2009;109:159-166